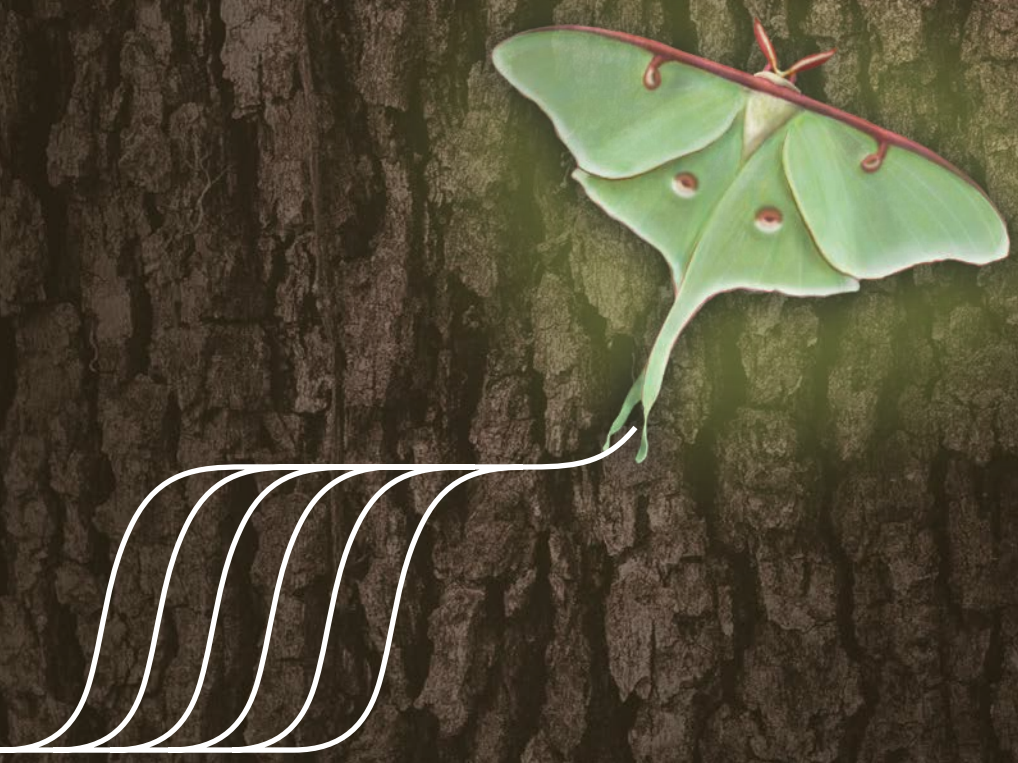
Issue | **2017**

NEB expressions

a scientific update



in this issue

NEW ENGLAND BioLabs® Inc.

be **INSPIRED**
drive **DISCOVERY**
stay **GENUINE**

FEATURE ARTICLE by Gregory C. Patton, Ph.D., Andrew N. Gray, Ph.D., Nathan A. Tanner, Ph.D., Janine G. Borgaro, Ph.D., Yan Xu, Ph.D., Julie F. Menin, M.S. & Nicole M. Nichols, Ph.D., New England Biolabs, Inc.

Development of a high-throughput data analysis method for quantitative real-time PCR (qPCR)

Over the last 20 years, quantitative real-time PCR (qPCR) has become an essential technique in molecular biology for detecting and quantifying nucleic acids. Workflow simplicity and advances in instrumentation now permit sizeable quantities of data to be generated rapidly, with 96, 384, or even 1536 reactions in one qPCR experiment. The challenge lies in the details: qPCR experiments require thoughtful design and analysis to capture all relevant information, such that accurate and appropriate conclusions can be drawn.

Development of NEB's Luna® qPCR product line required repeated data collection on a series of test panels, each containing multiple targets. It became clear during early development that a more scalable approach to data analysis and visualization was required to better understand how changes in reagent composition impacted performance. In order to compare various amplicon panels over multiple qPCR runs, instruments, reagents and conditions, a high-throughput data analysis method termed "dots in boxes" was developed. The output of this analysis captures key assay characteristics, highlighted in MIQE guidelines, as a single data point for each qPCR target. This method of analysis permits multiple targets and conditions to be compared in one graph, allowing concise visualization and rapid evaluation of overall experimental success.

INTRODUCTION TO qPCR

qPCR is a powerful fluorescence-based technique that detects and quantifies nucleic acids in a variety of samples. In 1992, Higuchi et al. showcased the first example of real-time PCR by using a camera during the amplification reaction to continuously monitor the incorporation of ethidium bromide, an intercalating dye that fluoresces in the presence of double-stranded DNA under ultra-violet light (1). Currently, most qPCR experiments commonly employ the dsDNA intercalating dye SYBR® Green I or hydrolysis probes (e.g., TaqMan®) to monitor amplification (2). Plotting the measured fluorescence signal versus PCR cycle number results in a graphical representation of amplification. The point at which the fluorescence signal exceeds the background fluorescence level is known as the quantification cycle (C_q). Comparing C_q values permits evaluation of relative target abundance between two or more samples. Alternatively, C_q values can be used to calculate absolute target quantities via reference to an appropriate standard curve, derived from a series of known DNA or RNA dilutions. This technique can be more powerful than traditional PCR, allowing both qualitative information (presence or absence of a target sequence), as well as the quantitative data (nucleic acid quantity) to be determined without opening the reaction tube. Greater sensitivity and lower risk of carryover contamination has resulted in qPCR replacing end-point PCR in many applications. Today, the technique is used in a variety of fields, from molecular diagnostics to agricultural research, and in applications including mutation detection, genotyping, copy number variation and gene expression analysis.

MIQE GUIDELINES

Rapid adoption of qPCR and its relatively straightforward execution (mixing amplification reagents, primers and template) has led to the generation of an enormous amount of data, as evidenced by the numerous publications containing qPCR experiments. However, the ease of generating qPCR data has also proven to be the technique's greatest challenge (3). A diverse set of protocols, instruments, reagents and analysis methods can be found in the scientific literature, with many publications reporting invalid or conflicting data sets. The lack of consensus on best experimental practices for qPCR resulted in the establishment of the Minimum Information for Publication of Quantitative Real-Time PCR Experiments (MIQE) guidelines by Bustin et al. (4).

The MIQE guidelines established a set of qPCR performance metrics that should be determined and reported in peer-reviewed publications to ensure robust assay performance and reproducibility.

These assay characteristics include:

- PCR efficiency
- Linear dynamic range
- Limit of detection (LOD)
- Target specificity
- Precision

One of the most important assay characteristics is PCR efficiency, which is a measure of product duplication at every amplification cycle. PCR efficiency is measured by amplifying multiple known concentrations of nucleic acid to obtain C_q values for each concentration. A standard curve is created by plotting the observed C_q values on the y-axis and the $\log_{10}$ of the template concentration on the x-axis. Efficiency is calculated using the equation:

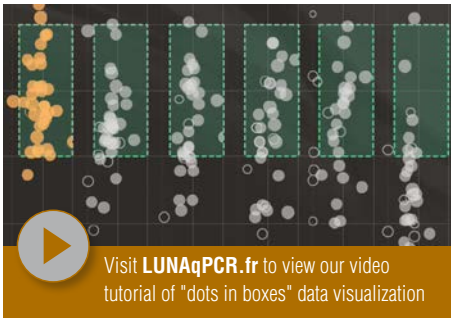
PCR efficiency = $10^{-1/\text{slope}} - 1$. A slope of -3.32 represents 100% PCR efficiency and indicates doubling of the target amplicon at each PCR cycle.

The linear dynamic range establishes the upper and lower limits for quantification and should be linear for at least three log10 concentrations of template. Preferably, the dynamic range encompasses five to six orders of magnitude. Linearity over a dynamic range is reported by the R^2 coefficient of determination for the C_q values linear fit to the standard curve.

The limit of detection is often defined as the lowest concentration at which 95% of target sequences are detected in positive samples. An ideal Poisson distribution and single copy detection dictate the lowest theoretical LOD is 3 molecules per PCR. Its determination establishes the lower boundary for target detection with 95% confidence (5).

Target specificity should be confirmed by product size, sequencing or melt curve analysis, since primers may unexpectedly amplify off-target regions. In addition, some primer sets have a propensity to form primer dimers during amplification, resulting in inaccurate quantification or false positive results. In order to identify spurious amplification products, no-template controls (NTC) should be included in every qPCR run. As NTCs can identify both unintended amplification products as well as contamination, criteria should be established for using these controls to determine when data should be accepted or rejected.

The last factor that should be evaluated is assay precision. Multiple replicates of the same sample should typically have high concordance. Variation inherently increases as the copy number decreases, but also can be attributed to factors such as pipetting errors and instrumentation.



DOTS IN BOXES ANALYSIS OF qPCR DATA

The MIQE-highlighted described above served as a guide for evaluating reagent performance during development of NEB's new Luna qPCR and RT-qPCR product line. To ensure strong performance across a range of amplicons, multiple test panels were created, with each panel containing a minimum of five targets that could be run in 96 or 384-well formats. Panels comprised of gDNA and cDNA targets were used to evaluate DNA-based qPCR master mixes, whereas RNA targets of varying abundance were used to assess RT-qPCR reagents. In general, targets spanned typical qPCR amplicon lengths (~70 to 200 bp), as well as GC content (~40 to 60%). Given the large data set that was created during development, data mining to decipher what changes impacted performance became challenging, and it was clear that a better, more scalable approach to data visualization was needed.

The fundamental performance criteria outlined in the MIQE publication therefore served as a basis for the development of a high-throughput data analysis method termed "dots in boxes" (Figure 1). For each amplicon, PCR efficiency, linear dynamic range, target specificity and precision was captured as a single data point plotted in two dimensions, with the PCR efficiency plotted on the y-axis and the delta C_q (ΔC_q) as the x-axis. ΔC_q is the difference between the C_q values of the lowest template dilution and the NTC. Setting guidelines around the typical accepted values for these two plotted parameters (PCR efficiency of 90 to 110% and ΔC_q of 3 or greater) created a graphical box, highlighting where successful qPCR experiments (dots) should fall.

TABLE 1:
Criteria for developing quality score metrics for "dots in boxes" analysis

	Intercalating Dye Chemistry	Hydrolysis Probe Chemistry
Linearity	$R^2 \geq 0.98$	$R^2 \geq 0.98$
Reproducibility	Replicate curves shall not vary by more than 1 C_q.	Replicate curves shall not vary by more than 1 C_q.
RFU Consistency	Maximum endpoint fluorescence signal for all curves shall be within 20% of the mean. Fluorescence signal shall not be jagged.	Increase of fluorescence signal shall be consistent for all curves, exhibiting parallel slopes. Fluorescence signal shall not be jagged.
Curve Steepness	Curves shall rise from baseline to plateau within 10 C_q values or less.	Curves shall rise from baseline to 50% maximum RFU within 10 C_q values or less.
Curve Shape	Curves shall exhibit a sigmoidal shape, resulting in a plateau of fluorescence signal.	Curves need not be sigmoidal, but shall appear to be reaching a horizontal asymptote by the last PCR cycle.

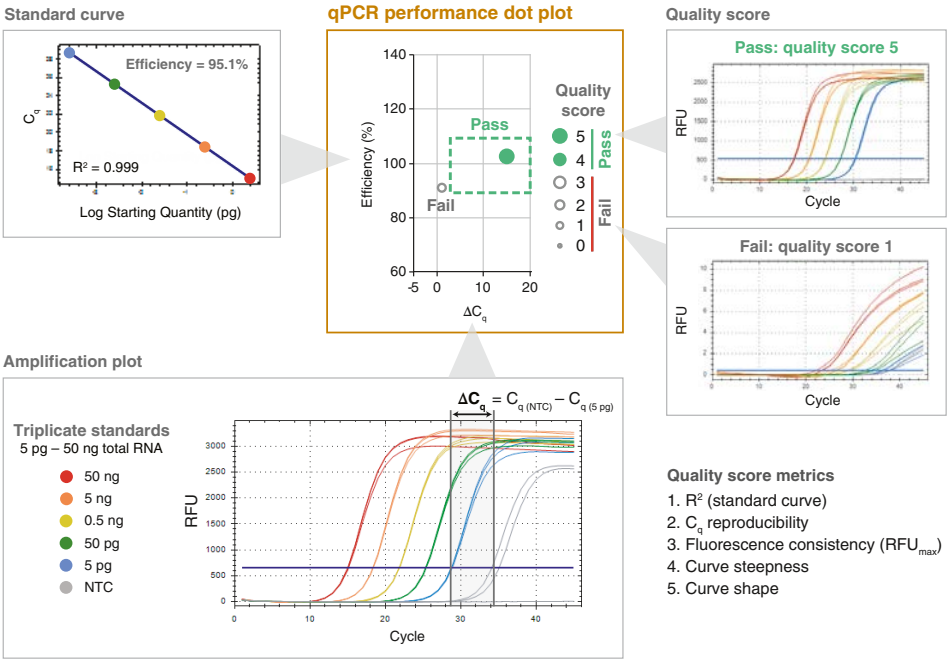
While this simple dot plot was informative on its own, it wasn't sufficient to capture all of the relevant details of each qPCR experiment. In order to represent additional information, such as the linearity of the dynamic range (R^2), the overall quality of the qPCR data was scored on a scale of 1 to 5, with 5 representing the highest quality. This scoring method was built upon previous work by Hall et al. (6). Additional performance criteria captured using the 5-point quality score included precision (reproducibility), fluorescence signal consistency, curve steepness and sigmoidal curve shape. Parameters for these five criteria were established to identify when the quality score should be penalized. Scoring criteria differed slightly for probe-based chemistry compared to intercalating dye-based detection (Table 1) due to differences in typical curve shape.

Once assigned, the quality score for each amplicon was represented by the dot size and opacity. The higher the quality score, the larger the dot. Additionally, quality scores of 4 and 5 were represented as solid dots while a score of 3 or less was captured as an open circle for simple visual screening of performance. Amplicons falling in the box and receiving a quality score of 4 or 5 represented high quality, reliable qPCR data. The dots in boxes method allowed multiple targets and conditions to be plotted on a single graph and compared quickly, creating an efficient, high-throughput visual method for data analysis.

To rigorously test qPCR performance, experiments were designed to simultaneously evaluate efficiency over a broad dynamic range of input concentrations; sensitivity by assessing low-input detection; and specificity by assessing off-target amplification. To accomplish this, qPCR efficiency was measured over a five-log dilution of template with data collected in triplicate for each dilution and a NTC. For genomic targets, an average of ~2 copies per

FIGURE 1:
Breaking it down: how we translate qPCR data into "dots in boxes"

NEB has developed a method to better evaluate the large amount of qPCR data generated in an experiment. The output of this analysis is known as a dot plot, and captures the key features of a successful, high-quality qPCR experiment as a single point. This method of analysis allows many targets and conditions to be compared in a single graph. For each experiment, triplicate reactions are set up across a five-log range of input template concentrations (Amplification plot, bottom-left). Three non-template control (NTC) reactions are also included, for a total of 18 reactions per condition/target. Efficiency (%) is calculated (Standard curve, top-left) and is plotted against ΔC_q (dot plot, center), which is the difference between the average C_q of the NTC and lowest template dilution. This parameter captures both detection of the lowest input and non-template amplification. Acceptable performance criteria are defined as an Efficiency of 90-110% and a ΔC_q of ≥ 3 (green box). Other performance criteria are captured using a 5-point Quality Score (top-right). Quality Score is represented by the size and fill of the plotted dot, with experiments that pass all performance criteria represented by a solid dot within the box.



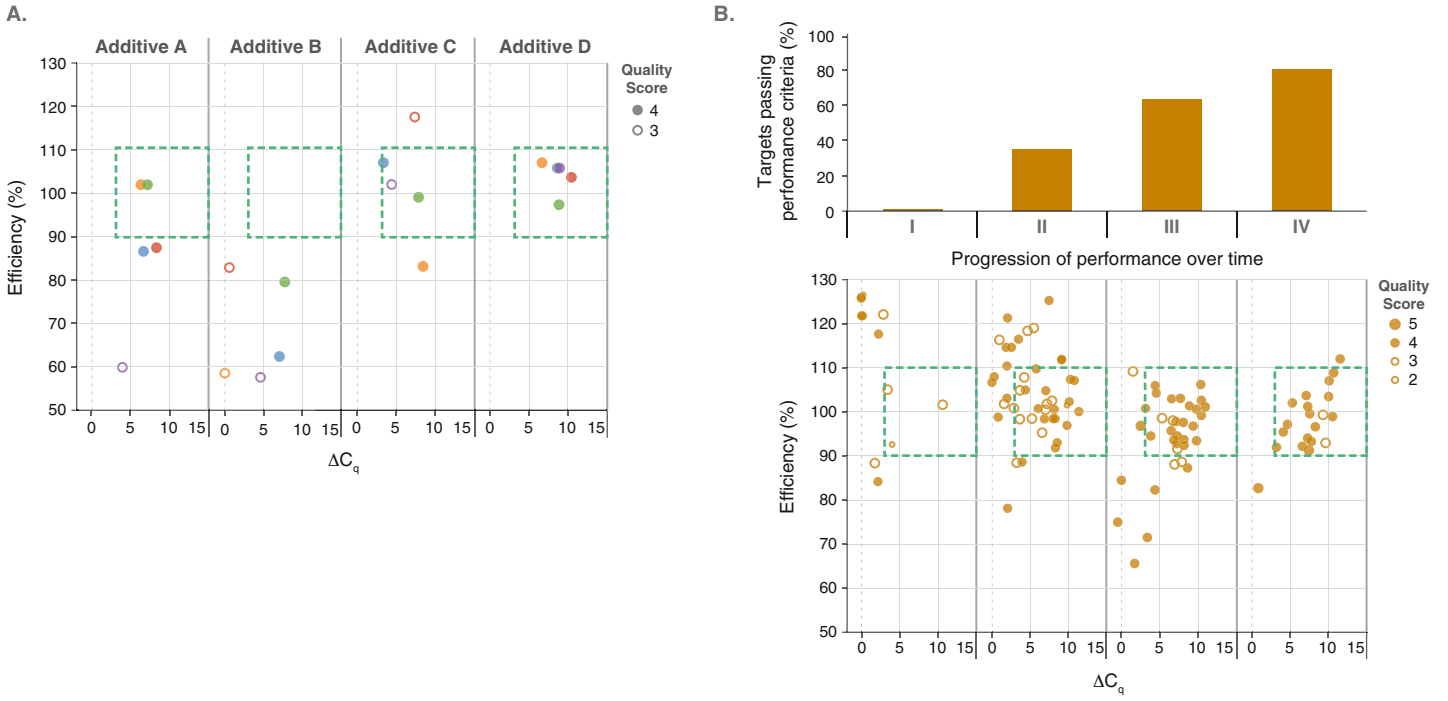
Quality score metrics

1. R^2 (standard curve)
2. C_q reproducibility
3. Fluorescence consistency (RFU_{max})
4. Curve steepness
5. Curve shape

continued on page 4...

FIGURE 2:
"Dots in boxes" enables visual screening of reagent optimization

The data analysis tool, dots in boxes, was used during development to improve qPCR reagent performance. Data was collected for qPCR targets varying in length and GC content, using Jurkat genomic DNA as input. Results were evaluated for efficiency, low input detection and lack of non-template amplification (where ΔC_q = average C_q of non-template control – average C_q of lowest input). In addition, consistency, reproducibility and overall curve quality were assessed (Quality Score, Table 1). A) Additives A through D were screened on five amplicons, each represented by a colored dot, to examine their effect on qPCR performance. Additive D resulted in successful amplification of all targets while Additive B was detrimental to amplification, resulting in low PCR efficiencies. B) Dots in boxes permitted large volumes of data to be compared over multiple master mix compositions, ultimately driving reagent optimization. Progression of performance is displayed for several predecessors of the Luna Universal qPCR Master Mix (NEB #M3003). Mixes with successful qPCR performance were built upon to establish the final composition of the Luna products.



reaction was routinely tested to assess the limits of low input detection. Since the ΔC_q incorporates both the C_q of the lowest input and that of the NTC ($\Delta C_q = C_{q(NTC)} - C_{q(lowest\ input)}$), it allows sensitivity and specificity to be captured in a single variable. Inability to amplify the lowest template dilution results in a ΔC_q of 0 in most cases, since curves failing to cross the threshold are automatically given a C_q value corresponding to the total number of amplification cycles. The presence of non-specific or contaminating amplification in NTC reactions also reduces the ΔC_q, such that either lack of low-input amplification or excessive off-target amplification can push the ΔC_q below the passing (≥ 3) threshold. Target specificity was also evaluated using denaturation or melt curves for all intercalating dye-based qPCR assays, although this information was not captured in the dot plot.

Pairing dots in boxes with an existing custom laboratory information management system (LIMS) permitted the performance of reagents to be screened and tracked on all amplicon panels. The LIMS, previously established for the development of NEB's Q5® High-Fidelity DNA Polymerase products, was modified to capture all relevant experimental details. The database connected results from each qPCR experiment (e.g., C_q values, PCR efficiency, and linearity) to the contents of each well in that experiment (e.g., target, template concentration, primer concentration, qPCR master mix, additives, etc.) such that performance could be linked to reaction variables and conditions. Additional details including the operator, real-time PCR instrument ID, and cycling conditions were also recorded. Tableau®, an analytics software package, was used to analyze the data and to create graphical displays of the dot plots. An example outcome is shown in Figure 2A. Here, the impact of known PCR additives and the concentration ranges that were beneficial to performance were quickly assessed on a development lot of the DNA dye-based master mix. Additive D resulted in the best performance on this particular panel of five amplicons. Unfortunately, improved performance on one particular qPCR panel did not necessary translate to positive performance across all panels evaluated. Thus, the development process was by necessity methodical and iterative. This made the ability to analyze and visualize large sets of results, covering multiple test panels, formulations and experimental conditions, all the more crucial. Dots in boxes thus played a critical role in the development of NEB's Luna products, driving reagent optimization by quickly identifying compositions with increased performance across all test panels. Successful compositions were built upon and fine-tuned, progressively improving the percentage of amplicons that fell in the box with high quality scores (Figure 2B), and thus overall performance. As a result, the final Luna qPCR formulations exhibit robust performance on diverse targets from a wide range sample types and sources.

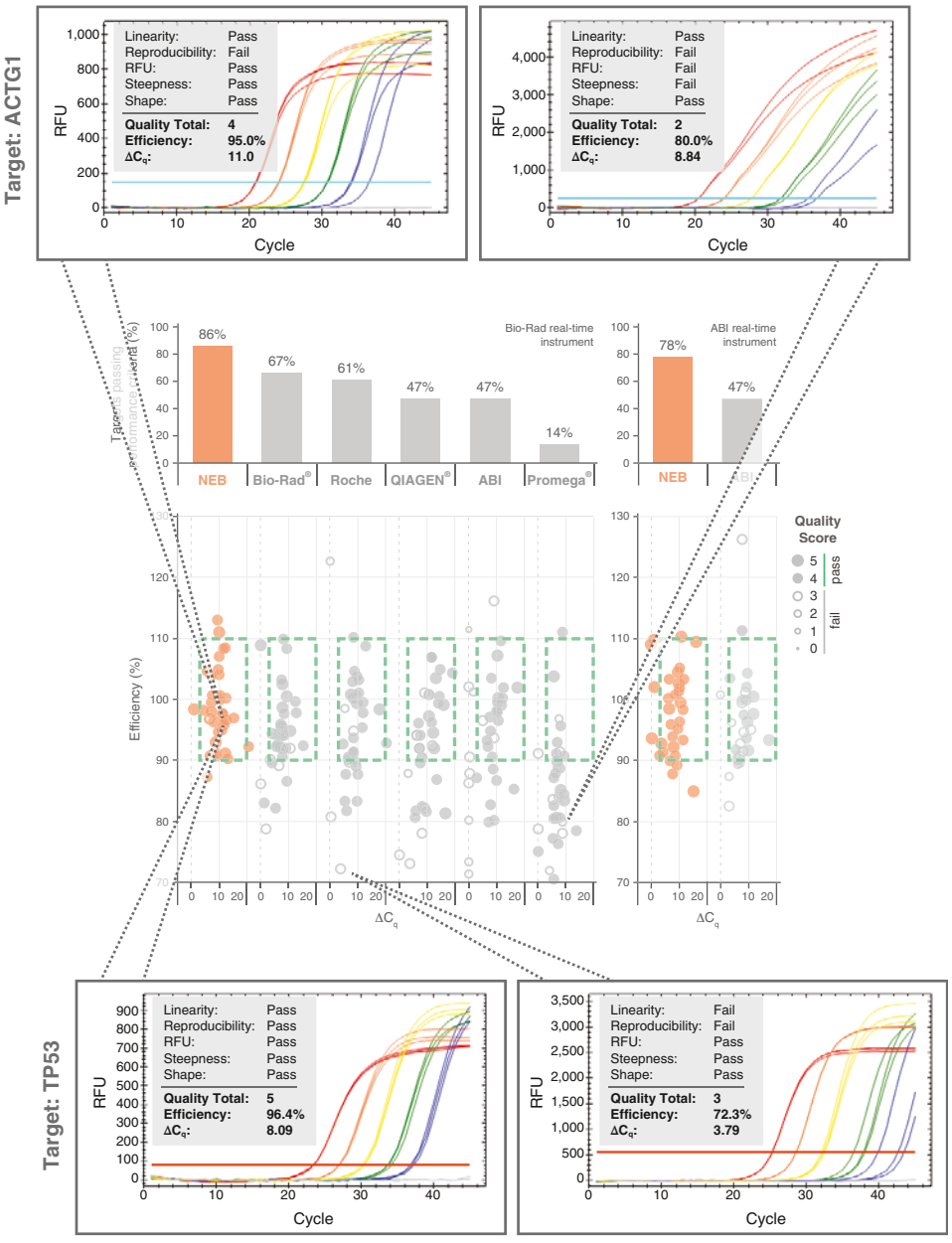
DOTS IN BOXES AS A COMPARISON TOOL

Dots in boxes also permitted large-scale performance comparisons between the Luna Universal qPCR and RT-qPCR reagents to various other commercial product offerings. Each commercial mix was challenged against test panels containing a range of targets. Amplicon panels used during the development were tested with a variety of commercial mixes and commercial primer/probe sets. Data was collected by two separate users and experiments were performed according to each manufacturer's specific product recommendations. The results for the Luna Universal qPCR Master Mix (NEB #M3003) are shown in Figure 3. Luna generates the highest quality qPCR data of all reagents tested, with 86% of all amplicons tested falling in the box with high quality scores. Strong performance was observed for the entire Luna portfolio; dots in boxes performance comparisons for each Luna product can be found at **www.lunaqpcr.fr**.

FIGURE 3:

Luna qPCR products outperform other commercially-available reagents

qPCR reagents from NEB and other manufacturers were tested across 16–18 qPCR targets varying in length and GC content, using either Jurkat genomic DNA or Jurkat-derived cDNA as input (10 genomic DNA targets and 8 cDNA targets on Bio-Rad real-time instrument, 9 genomic and 7 cDNA targets on ABI instrument). For each testing condition, data was collected by 2 users and according to manufacturers' specifications. Results were evaluated as described in Figure 2. Representative curves are shown for two targets, ACTG1 (top) and TP53 (bottom), to demonstrate the correlation of dots in boxes with typical qPCR data. Results for NEB and other major manufacturers are shown: Bio-Rad, SsoAdvanced™ Universal SYBR® Green Supermix; Roche, FastStart™ SYBR Green Master; QIAGEN®, QuantiTect® SYBR Green PCR Kit; ABI, PowerUP™ SYBR Green Master Mix; Promega®, GoTaq® qPCR Master Mix. Luna Universal qPCR Master Mix from NEB outperformed all other reagents tested.



CONCLUSION

Dots in boxes is a powerful, high-throughput data analysis method based on the MIQE guidelines that enables rapid, concise comparison of qPCR performance across many targets and for multiple reagents, conditions and/or protocols, permitting an overview of qPCR performance over thousands of reactions where such visualization was not previously possible. Combining the dots in boxes analysis method, a range of target test panels, and a custom LIMS enabled us to create and mine large data sets for information, identify critical variables that affect amplification in qPCR, and harness this information to optimize qPCR reagents. The dots in boxes analysis tool was thus invaluable in development of the Luna qPCR and RT-qPCR reagents, and will continue to benefit future qPCR evaluation and development efforts.

References

1. Higuchi, R., et al. (1992) *Biotechnology* 10, 413-417.
2. Navarro, E., et al. (2015) *Clin. Chim. Acta.* 439, 231-250.
3. Bustin, S.A., and Nolan, T. (2004) J. *Biomol Tech.* 3, 155-156.
4. Bustin, S.A., et al. (2009) *Clin. Chem.* 55, 611-622.
5. Wittwer, C.T., et al. (2004) *Washington: ASM Press.* 71-84.
6. Hall, A.T., et al. (2013) *PLOS One 8*: e73845.

BIO-RAD® is a registered trademark of Bio-Rad Laboratories, Inc.
SYBR® is a registered trademark of Molecular Probes, Inc.
QIAGEN® and QUANTITECT® are registered trademarks of Qiagen, Inc.
GOTAQ® and PROMEGA® are registered trademarks of Promega Corporation
TABLEAU® is a registered trademark of Tableau Software
FASTSTART™ is a trademark of Roche
POWERUP™ is a trademark of Thermo Fisher Scientific
SSOADVANCED™ is a trademark of Bio-Rad Laboratories

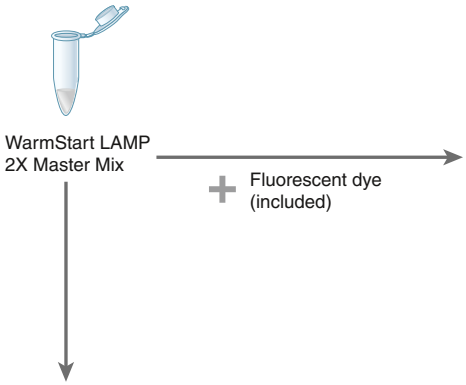
Learn more about NEBs new LUNA Universal qPCR & RT-qPCR Reagents on page 10.

New Products for Isothermal Amplification

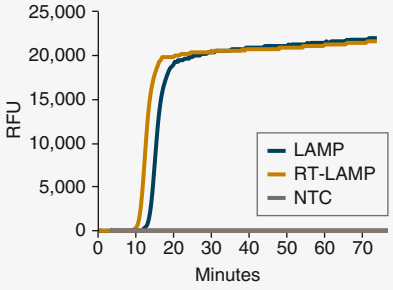
WarmStart® LAMP Kit (DNA & RNA)

Loop Mediated Isothermal Amplification (LAMP) is a commonly-used technique for rapid nucleic acid detection. NEB's WarmStart LAMP products provide a simple, one-step solution for DNA or RNA targets. The master mix supplied with the WarmStart LAMP Kit contains the robust and rapid Bst 2.0 WarmStart DNA Polymerase and WarmStart RTx Reverse Transcriptase, both *in silico*-designed enzymes for improved performance in LAMP reactions. The kit also includes a fluorescent dye to enable real-time fluorescence measurement of LAMP. The WarmStart LAMP Kit is compatible with multiple detection methods, as shown in the figure below.

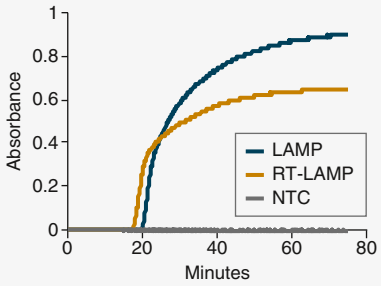
NEB's WarmStart LAMP Kit (DNA & RNA) is compatible with multiple detection methods*

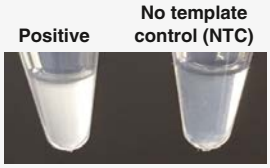


Real-time fluorescence measurement



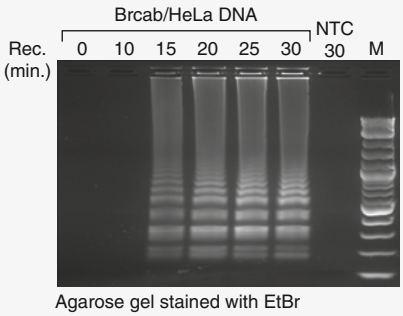
Turbidity detection





LAMP product is visible by eye due to the precipitation of magnesium pyrophosphate

Endpoint visualization



Agarose gel stained with EtBr

Visualization using a fluorescent intercalating dye (e.g., ethidium bromide, SYBR® green or calcein/HNBlue)

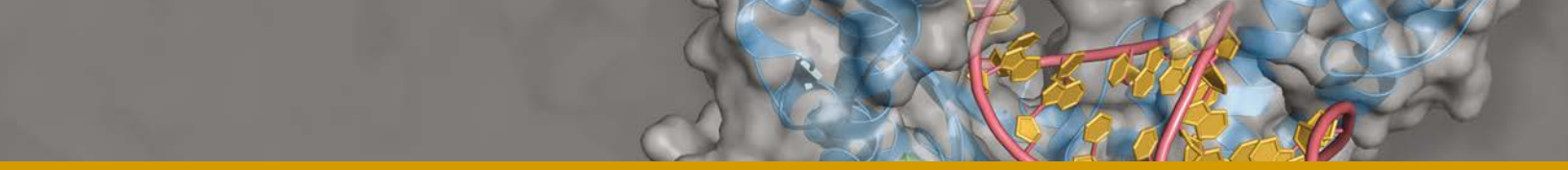
** The NEB WarmStart LAMP Kit (DNA & RNA) includes separate fluorescent dye for real-time fluorescence measurement. Alternately, detection can be accomplished by turbidity detection or endpoint visualization.*

looking for best-in-class visual detection?

Try our WarmStart Colorimetric LAMP 2X Master Mix (DNA & RNA)

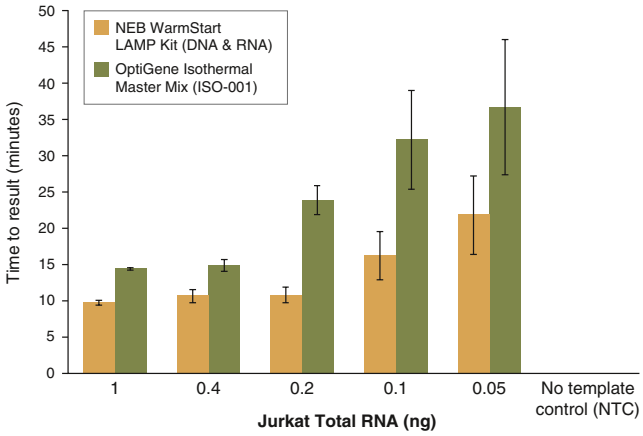
- NEB's WarmStart Colorimetric LAMP 2X Master Mix (DNA & RNA) offers the same robust performance as the WarmStart LAMP Kit, and contains a colorimetric dye for best in class visual detection of your target.

Yellow color indicates that amplification has occurred.



NEB's WarmStart LAMP Kit (DNA & RNA) offers speed and robust sensitivity

A RNA target (HMBS2) was amplified from Jurkat total RNA using the WarmStart LAMP Kit and OptiGene Master Mix (ISO-001). Reactions were performed at 65°C for 74 minutes on a real-time thermocycler (Bio-Rad® CFX96) in triplicate. Time to result is set as the time at which the fluorescence crossed a threshold of 10% of maximal fluorescence. NEB's WarmStart LAMP Kit resulted in faster and more sensitive detection as compared to the OptiGene Master Mix.



ORDERING INFORMATION:

PRODUCT	NEB #	SIZE
WarmStart LAMP KIT (DNA & RNA)	E1700S/L	1,600/8,000 units
WarmStart Colorimetric LAMP 2X Master Mix (DNA & RNA)	M1800S/L	100/500 rxns
Bst 2.0 WarmStart DNA Polymerase	M0538S/M/L	1,600/8,000 units
WarmStart RTx Reverse Transcriptase	M0380S/L	50/250 rxns

optimization tips for LAMP

- Use LAMP primer design software (e.g., Primer Explorer – primerexplorer.jp/e/). Select 2–3 sets for each target and compare performance in a LAMP assay.
- Include loop primers for faster reactions
- Use high magnesium (6–8 mM) and dNTP (1–1.4 mM) concentrations for best results
- Omit betaine, unless it has a demonstrated benefit
- Optimize the reaction temperature (60–65°C for Bst LF and 63–70°C for Bst 2.0/3.0)
- To prevent contamination, use Bst 3.0 or Antarctic Thermo-labile UDG (NEB #M0372), which denatures rapidly

Visit www.neb-online.fr for more detailed information & to request the free brochure.

Did you know?

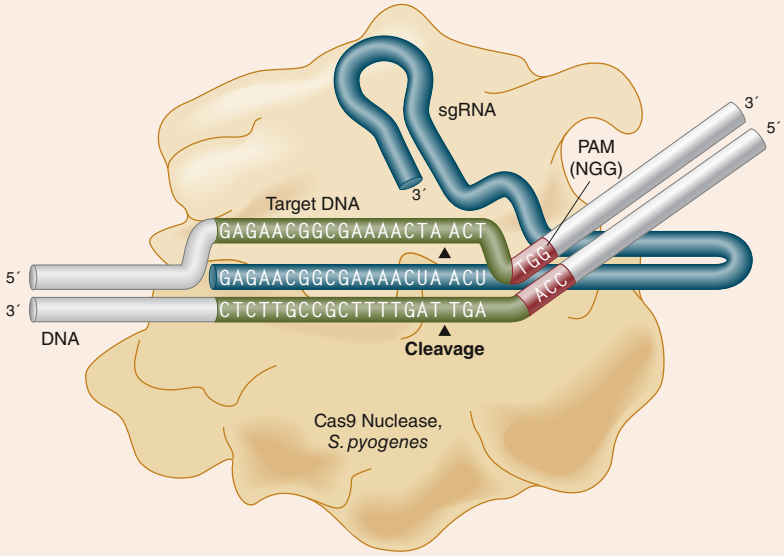
All of the wastewater from our Ipswich, MA campus is treated by a state-of-the-art Solar Aquatics® System. This system utilizes and accelerates the natural purification processes found in streams and wetlands, purifying the water for reuse or for groundwater recharge.

Learn more and take a tour of the facility at www.neb.com/environmentalphilosophy

Tools to support your CRISPR/Cas9

New England Biolabs provides reagents to support a broad variety of CRISPR/Cas9 genome editing approaches. From introduction of Cas9 and single guide RNA (sgRNA) on plasmids, to direct introduction of Cas9 ribonucleoprotein (RNP) and detection of edits using next generation sequencing or enzymatic mutation detection, NEB provides reagents that simplify and shorten genome editing workflows. Generating RNPs for direct introduction requires Cas9 protein and either sgRNA or separate crRNA and tracrRNA. EnGen® Cas9 NLS, *S. pyogenes* is engineered for high genome editing efficiency. The EnGen sgRNA Synthesis Kit combines template assembly and *in vitro* transcription for rapid generation of microgram quantities of custom sgRNA, requiring only a user-supplied single ssDNA oligonucleotide. To determine editing efficiency, the EnGen Mutation Detection Kit provides a full workflow from PCR amplification to T7 Endonuclease I-based mutation detection. Alternatively, NEB supplies Cas9 wild type and restriction enzymes, both of which can be used *in vitro* to determine the extent of editing.

Schematic representation of Cas9 Nuclease, *S. pyogenes* sequence recognition and DNA cleavage



Visit **www.neb.com/GenomeEditing** to view our online tutorial which walks through genome editing with Cas9

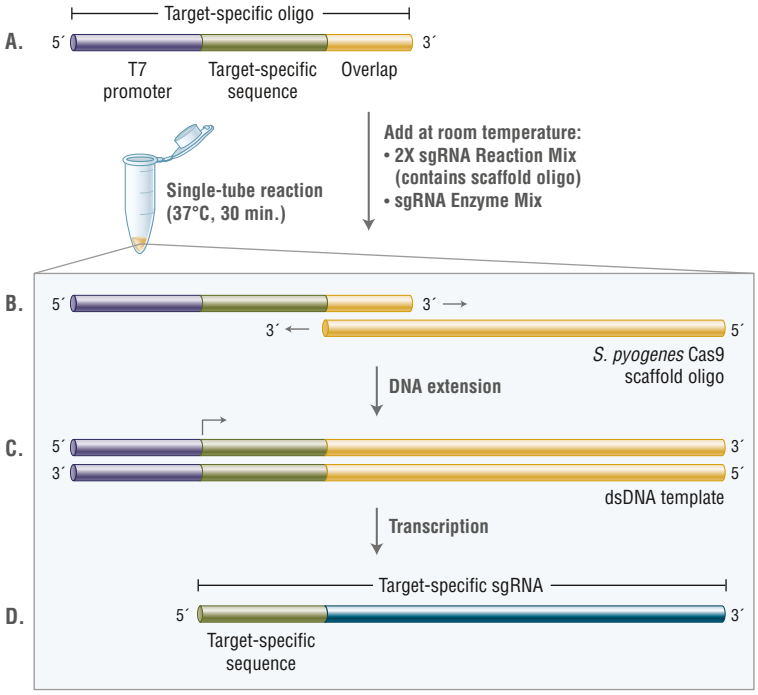
EnGen sgRNA Synthesis Kit

The EnGen sgRNA Synthesis Kit simplifies the generation of microgram quantities of custom sgRNAs in an hour or less by combining template synthesis and transcription. The single-tube reaction is easy to set up and requires a single ~55 nt ssDNA target-specific oligonucleotide, which is combined with the Reaction Mix and Enzyme Mix included in the kit. sgRNAs are suitable for use in downstream applications, including CRISPR/Cas9-based genome editing and *in vitro* DNA cleavage. This single-reaction format offers ease-of-use and eliminates separate DNA amplification and template clean up steps.

"This kit is really easy to use and will save us plenty of time in making sgRNAs! Thanks for the streamlined method!"

– Postdoctoral Researcher, Harvard University

EnGen sgRNA Synthesis Kit overview



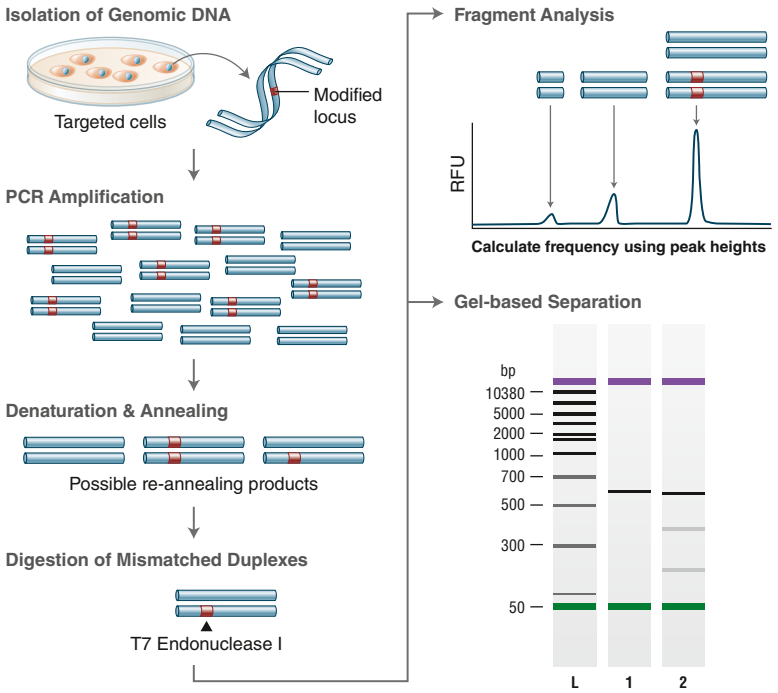
Genome Editing Workflows

EnGen Mutation Detection Kit

A widely used method to identify mutations is the T7 Endonuclease I mutation detection assay. This assay detects heteroduplex DNA that results from the annealing of a DNA strand, including desired mutations, with a wild-type DNA strand. The EnGen Mutation Detection Kit provides optimized reagents for performing robust T7 Endonuclease-based detection of genome editing events.

Workflow for EnGen Mutation Detection Kit

Genomic DNA is amplified with primers bracketing the modified locus. PCR products are then denatured and re-annealed yielding three classes of possible structures. Duplexes containing a mismatch greater than one base are digested by T7 Endonuclease I. The DNA is then electrophoretically separated and fragment analysis is used to estimate targeting efficiency.



ORDERING INFORMATION:

PRODUCT	NEB #	SIZE
EnGen Cas9 NLS, *S. pyogenes*	M0646T/M	400/2,000 pmol
EnGen sgRNA Synthesis Kit, *S. pyogenes*	E3322S	20 rxns
EnGen Mutation Detection Kit	E3321S	25 rxns
T7 Endonuclease I	M0302S/L	250/1,250 units
Q5® Hot Start High-Fidelity 2X Master Mix	M0494S/L	100/500 rxns

online resources

Plasmid Repositories
- addgene.org

CRISPR-gRNA Design Tools
- deskgen.com
- crispr.mit.edu
- zifit.partners.org/ZiFiT
- e-crisp.org
- chopchop.rc.fas.harvard.edu
- benchling.com

Online Forums
- groups.google.com/forum/#!forum/crispr

Organism-specific Resources
- wormcas9hr.weebly.com
- flyrnai.org

Visit www.neb.com/GenomeEditing to find:

Protocols for applications such as sgRNA synthesis and direct introduction of sgRNA/Cas9 complexes

EnGen sgRNA Template Oligo Designer (accessible through NEBioCalculator® at **NEBiocalculator.neb.com**)

Our latest **Genome Editing Brochure** available for download

Visit www.neb-online.fr for more detailed information & to request the free brochure.

LUNA®

Universal qPCR & RT-qPCR Reagents

Lighting the way.

Fluorescence-based quantitative real-time qPCR (qPCR) is the "gold standard" for the detection and quantitation of nucleic acids due to its sensitivity and specificity. Luna products from NEB are optimized for qPCR or RT-qPCR, and are available for either intercalating dye or probe-based detection methods. All Luna products provide robust performance on diverse sample sources and target types.

Each Hot Start Taq-based Luna qPCR master mix has been formulated with a unique passive reference dye that is compatible across a wide variety of instrument platforms, including those that require a ROX reference signal. This means that no additional components are required to ensure machine compatibility. The mixes also contain dUTP, enabling carryover prevention when reactions are treated with NEB's Antarctic Thermolabile UDG (NEB #M0372).

A blue visible dye assists in tracking the reagents when pipetting into clear, multi-welled PCR plates.

These features, combined with rapid, sensitive and precise real-time qPCR performance, make Luna the universal choice for all your qPCR experiments.

EXPERIENCE BEST-IN-CLASS PERFORMANCE

- All Luna products have undergone rigorous testing to optimize specificity, sensitivity, accuracy and reproducibility.
- Products perform consistently across a wide variety of sample sources.
- A comprehensive evaluation of commercially-available qPCR and RT-qPCR reagents demonstrates superior performance of Luna products.

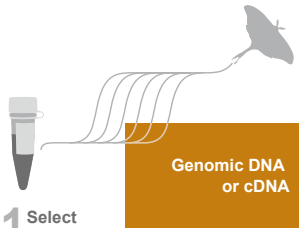
OPTIMIZE YOUR RT-qPCR WITH LUNA WARMSTART® REVERSE TRANSCRIPTASE

- Novel, thermostable reverse transcriptase (RT) improves performance.
- WarmStart RT paired with Hot Start Taq increases reaction specificity and robustness.

MAKE A SIMPLER CHOICE

- One product per application simplifies selection.
- Convenient master mix formats and user-friendly protocols simplify reaction setup.
- Non-interfering, visible tracking dye helps to eliminate pipetting errors.

Find the right Luna product for your application



	Dye-based	Probe-based
Genomic DNA or cDNA	Luna Universal qPCR Master Mix (NEB #M3003)	Luna Universal Probe qPCR Master Mix (NEB #M3004)
RNA	Luna Universal One-Step RT-qPCR Kit (NEB #E3005)	Luna Universal Probe One-Step RT-qPCR Kit (NEB #E3006)

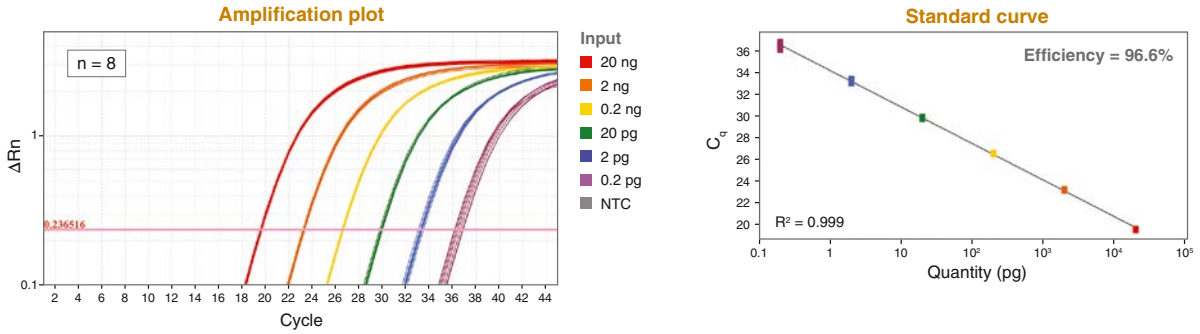
ORDERING INFORMATION

PRODUCT	NEB #	SIZE
Luna® Universal qPCR Master Mix	M3003S/L	200/500 rxns
Luna® Universal Probe Master Mix	M3004S/L	200/500 rxns
Luna® Universal One-Step RT-qPCR Kit	E3005S/L	200/500 rxns
Luna® Universal Probe One-Step RT-qPCR Kit	E3006S/L	200/500 rxns

Experience best-in-class performance for your qPCR & RT-qPCR

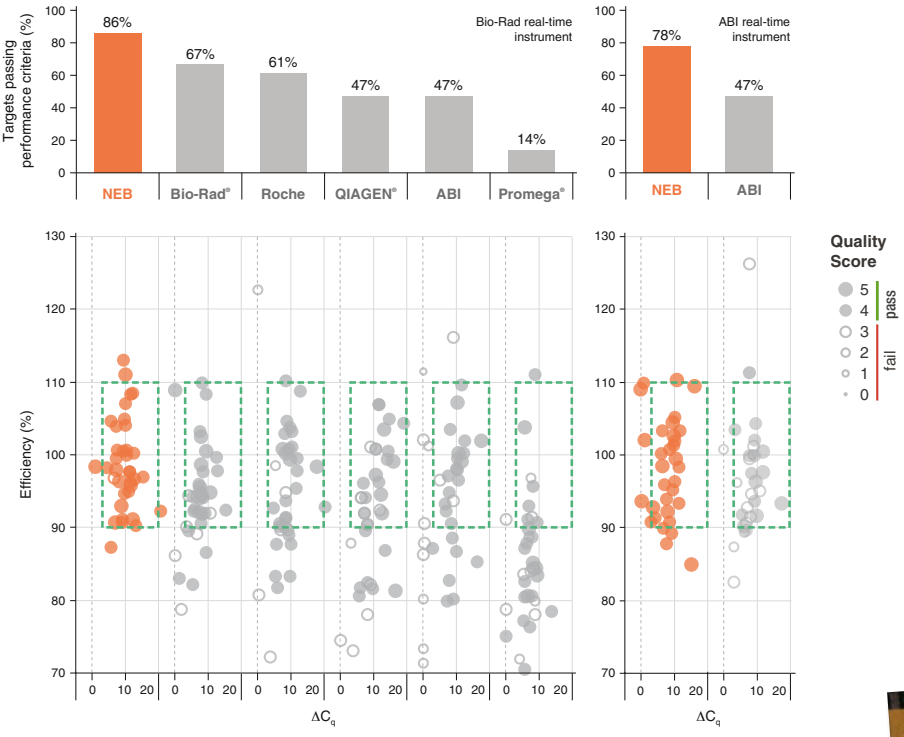
All NEB products undergo rigorous testing to ensure optimal performance, and Luna is no exception. We took into consideration numerous important traits when evaluating qPCR, including specificity, sensitivity, accuracy and reproducibility, to develop best-in-class qPCR reagents. Furthermore, we did a comprehensive evaluation of commercially-available qPCR and RT-qPCR reagents, and developed a method of analysis that allows you to quickly compare and evaluate the performance of these products. We wanted to be sure that Luna products will perform to your expectations for all your targets.

Luna products offer exceptional sensitivity, reproducibility and qPCR performance

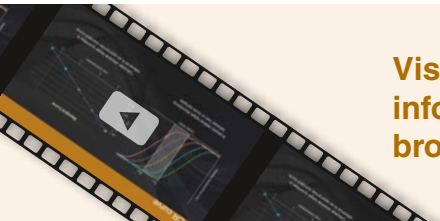


qPCR targeting human GAPDH was performed using the Luna Universal Probe qPCR Master Mix over a 6-log range of input template concentrations (20 ng – 0.2 pg Jurkat-derived cDNA) with 8 replicates at each concentration. cDNA was generated from Jurkat total RNA using the NEB Protoscript® II First Strand cDNA Synthesis Kit (NEB #E6560). NTC = non-template control

Evaluation of commercially-available dye-based qPCR reagents demonstrates the robustness and specificity of Luna



qPCR reagents from NEB and other manufacturers were tested across 16–18 qPCR targets varying in abundance, length and %GC, using either Jurkat genomic DNA or Jurkat-derived cDNA as input (10 genomic DNA targets and 8 cDNA targets on a Bio-Rad real-time instrument, 9 genomic and 7 cDNA targets on an ABI instrument). For each testing condition, data was collected by 2 users and according to manufacturer's specifications. Results were evaluated for efficiency, low input detection and lack of non-template amplification (where ΔC_q = average C_q of lowest input – average C_q of non-template control). In addition, consistency, reproducibility and overall curve quality were assessed (Quality Score). Bar graph indicates % of targets that met acceptable performance criteria (indicated by green box on dot plot and Quality Score > 3). Results for NEB and other major manufacturers are shown: Bio-Rad, SsoAdvanced™ Universal SYBR® Green Supermix; Roche, FastStart™ SYBR Green Master; QIAGEN, QuantiTect® SYBR Green PCR Kit; ABI, PowerUP™ SYBR Green Master Mix; Promega, GoTaq® qPCR Master Mix. NEB's Luna Universal qPCR Master Mix outperformed all other reagents tested.



Visit www.LUNAqPCR.fr for further information, free samples, videos & brochure request.

Request free LUNA® samples on next page

NEW ENGLAND BioLabs Inc.

New England Biolabs, Inc., 240 County Road, Ipswich, MA 01938-2723

GET A FREE T-SHIRT*

Visit **www.neb-online.fr** to request your free **LUNA® qPCR** or **RT-qPCR sample**, share your feedback and get a **free LUNA T-Shirt!** *

* Please visit **www.LUNAqPCR.fr** for details. Samples are limited to 2 per work group or as long as stock lasts.

COMING SOON!

The new Catalog & Technical Reference 2017/18

Request your free personal copy and update your address details!

France
New England Biolabs France SAS
Genopole Campus 1, Bâtiment 6
5 rue Henri Desbruères
91030 EVRY cedex
Toll Free Telephone: 0800 100 632
Toll Free FAX : 0800 100 610
info.fr@neb.com
www.neb-online.fr

USA (Headquarter)
New England Biolabs, Inc.
Telephone: (978) 927-5054
Toll Free: (U.S. Orders) 1-800-632-5227
Toll Free: (U.S. Tech) 1-800-632-7799
Fax: (978) 921-1350
info@neb.com

United Kingdom
New England Biolabs (UK), Ltd.
Call Free 0800 318486
info.uk@neb.com

Germany & Austria
New England Biolabs GmbH
Free Call 0800/246 5227 (Germany)
Free Call 00800/246 52277 (Austria)
info.de@neb.com

Canada
New England Biolabs, Ltd.
Toll Free: 1-800-387-1095
info.ca@neb.com

China, People's Republic
New England Biolabs (Beijing), Ltd.
Telephone: 010-82378265/82378266
info@neb-china.com

Japan
New England Biolabs Japan, Inc.
Telephone: +81 (0)3 5669 6191
info@neb-japan.com

Singapore
New England Biolabs Pte. Ltd.
Telephone +65 6776 0903
sales.sg@neb.com

www.neb-online.fr